Mol Bio 2: lectures 4 and 5

Sequence alignment

Substitution matrices

Multiple sequence alignment

BLAST

How sequences evolve

•point mutations (single base changes)

•deletion (loss of residues within the sequence)

•insertion (gain of residue within the sequence)

•truncation (loss of either end)

•extension (gain of residues at either end)

Mechanisms of insertion or extension:

- •duplication or whole gene or domain
- •polymerase "stutter"
- •transposable element
- •more??

How evolution is measured

point mutations substitution matrix score
insertion/deletion gap penalty
truncation/extension end gap penalty

Yes, an **alignment algorithm** is really A Model for Sequence Evolution!

 That means the way we do alignment should be closely aligned to what we know about how things evolve.

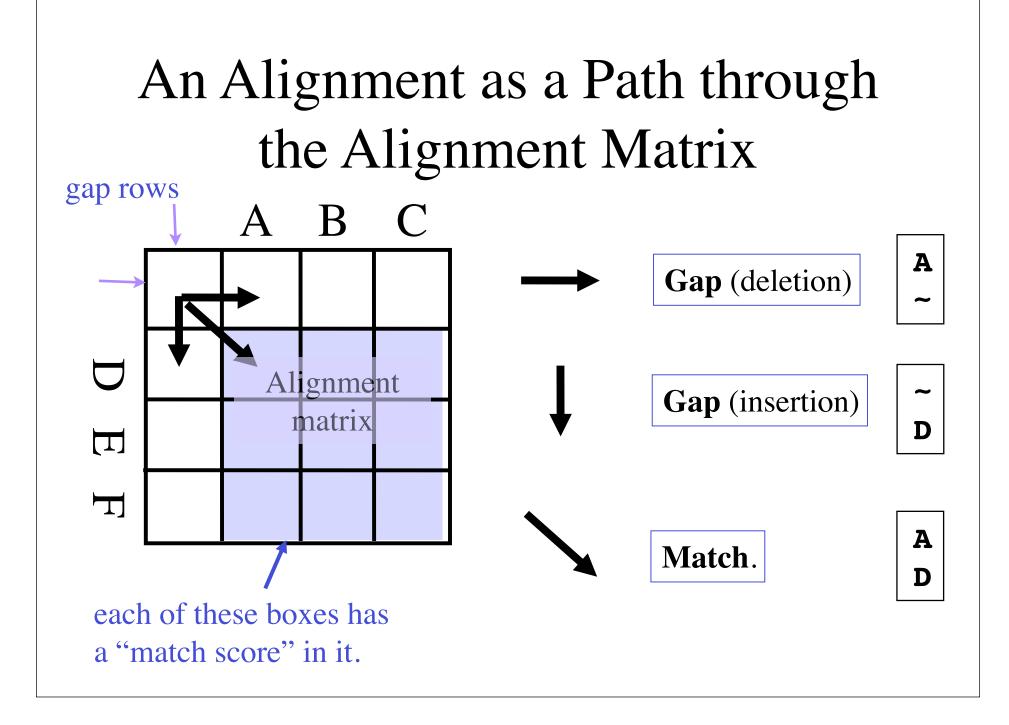
•point mutations relatively frequent, usually bad

•deletion infrequent, always bad, location dependent

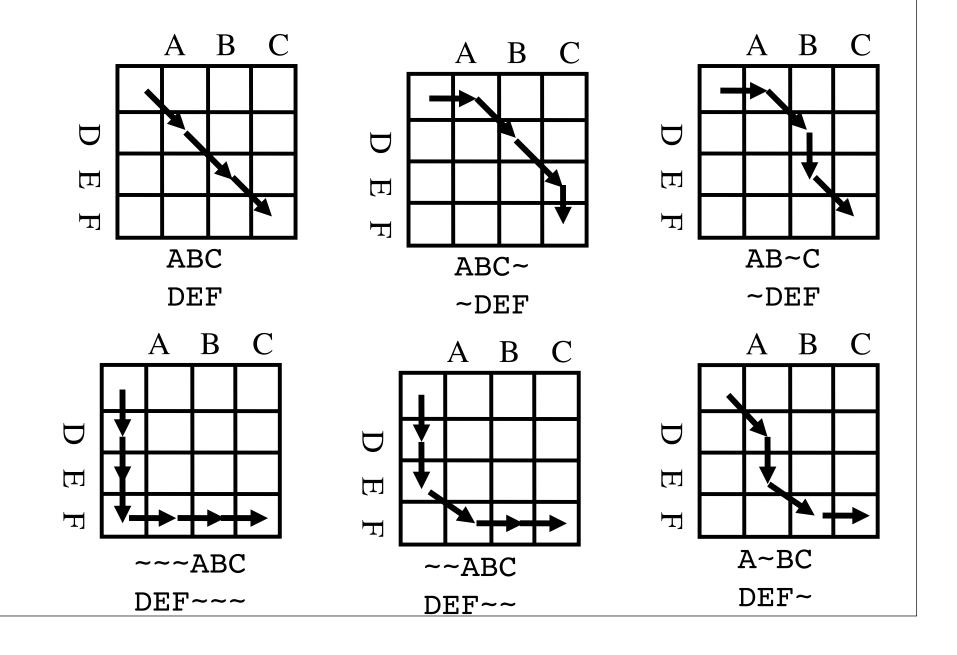
•insertion infrequent, always bad, location dependent

•truncation frequent, not so bad

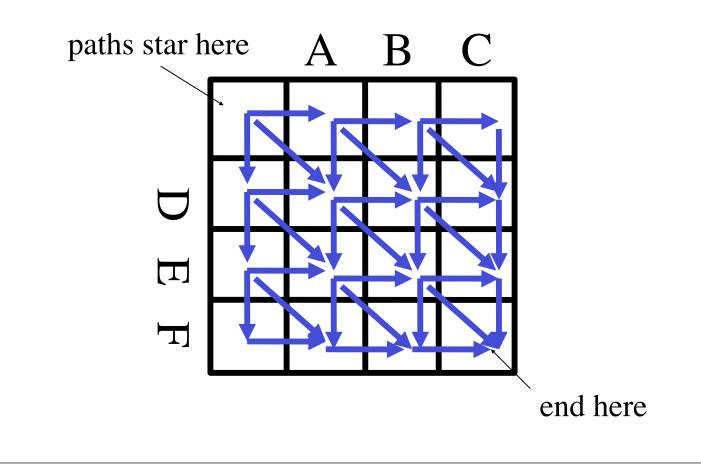
•extension frequent, not so bad



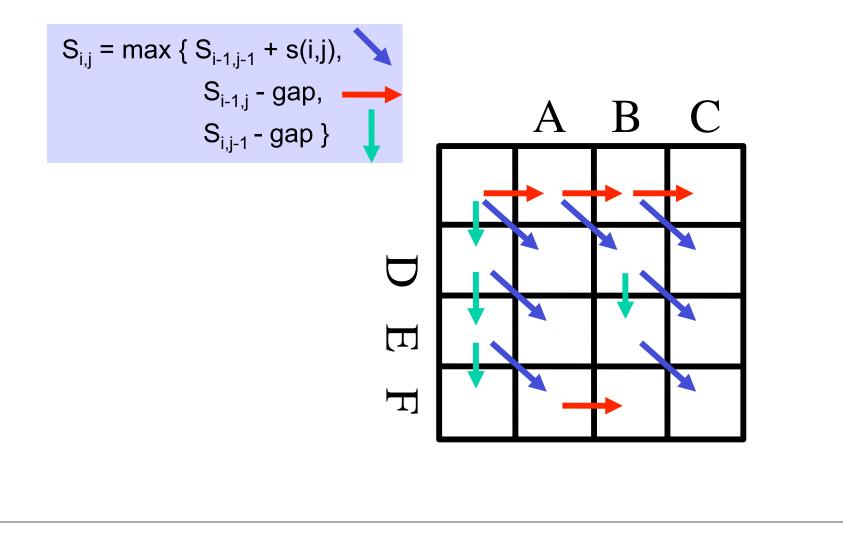
A walk through the alignment matrix



All possible arrow paths = all possible alignments





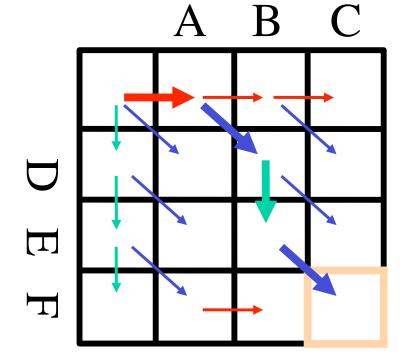


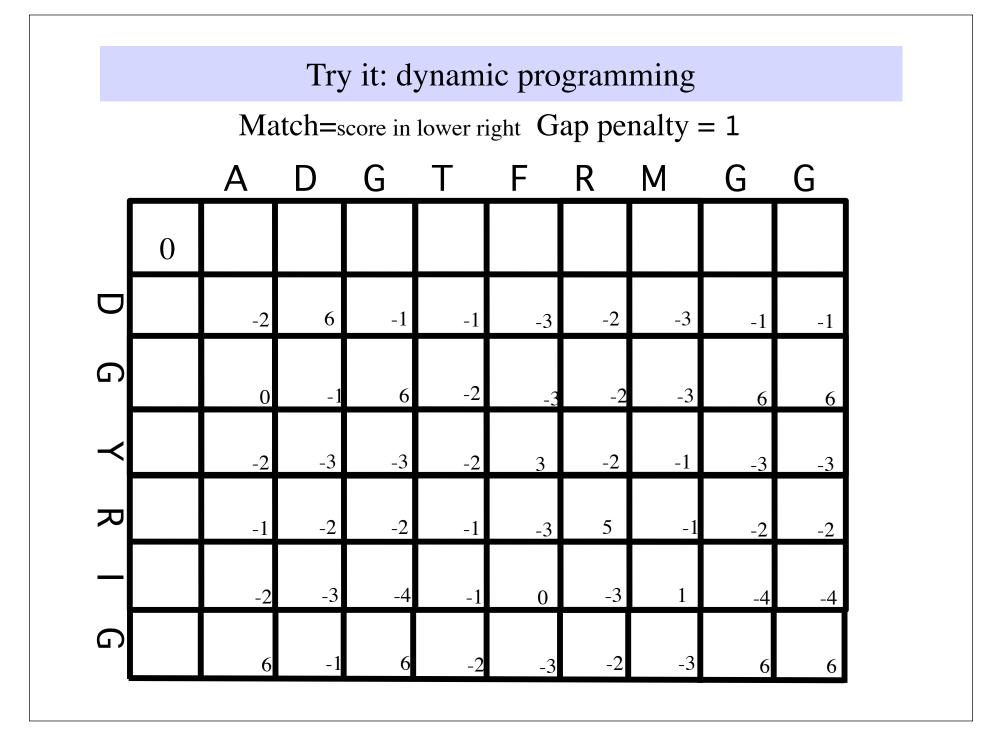
Traceback

Step 2: Trace arrows back to start.

Step 3: Alignment is constructed from the traceback arrows. Traceback starts from the **last box**

→ \ \ AB~C ~DEF



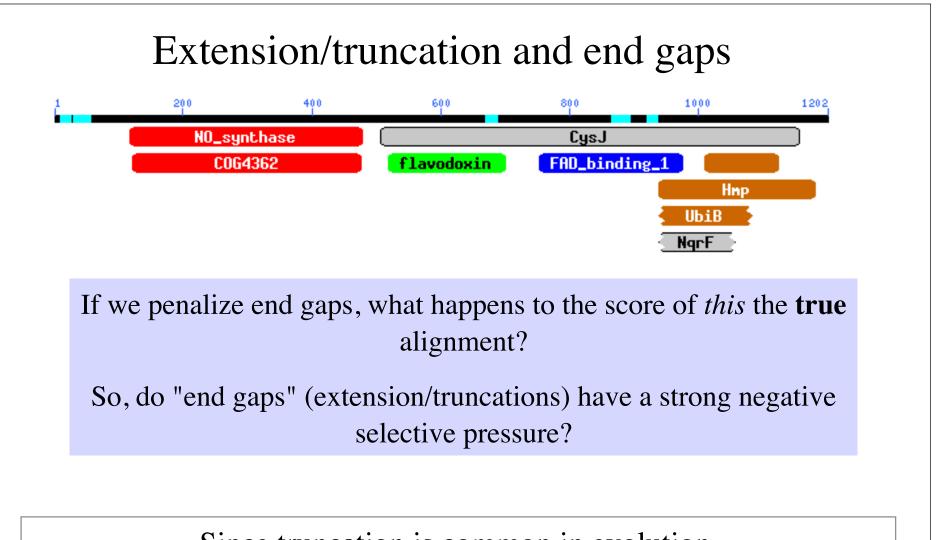


Does gap-to-gap make sense???

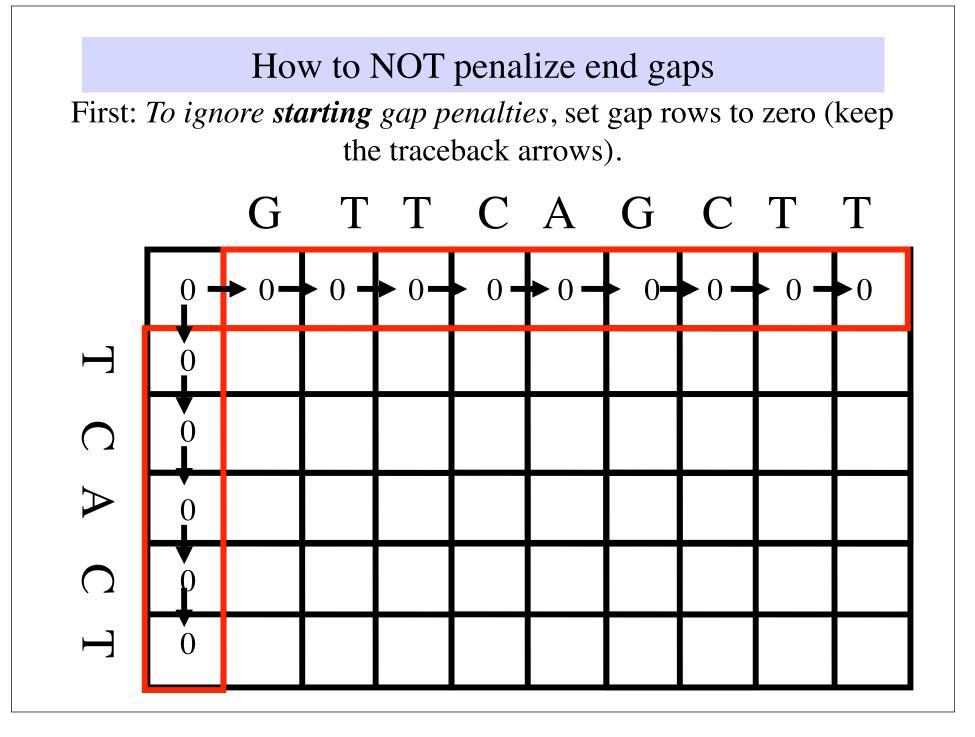
Special rules may apply for going directly from *insertion to* <u>deletion arrow</u>.

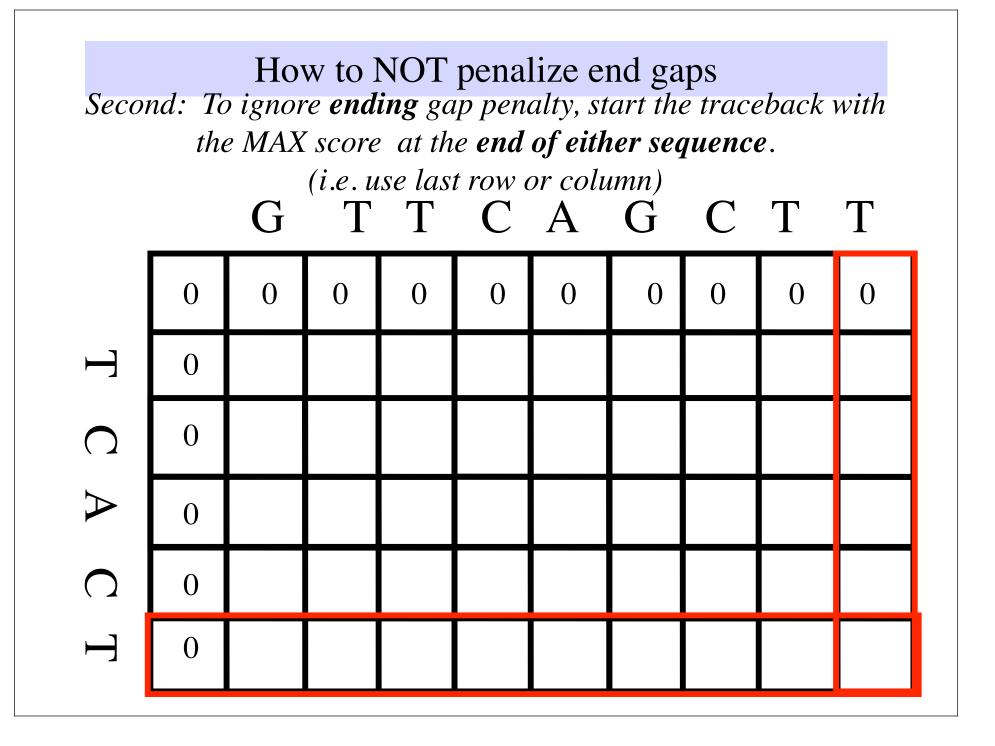
AGGCTACT~TATCA GGCTACTA~ATCA

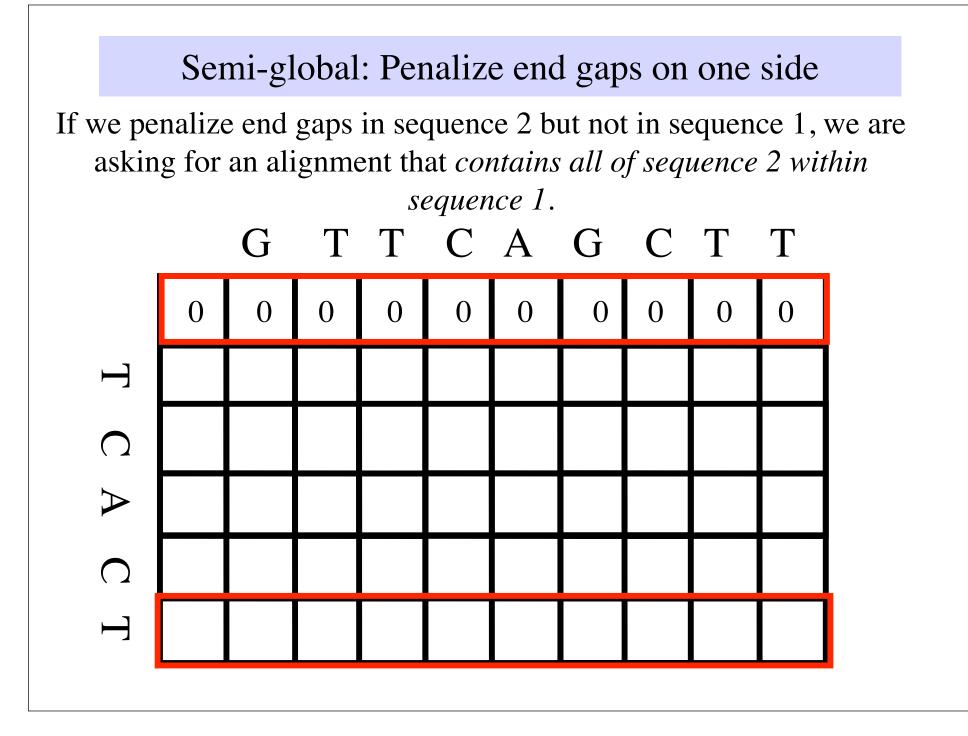
I to D can simply be **disallowed** in the DP algorithm. Most programs do this.

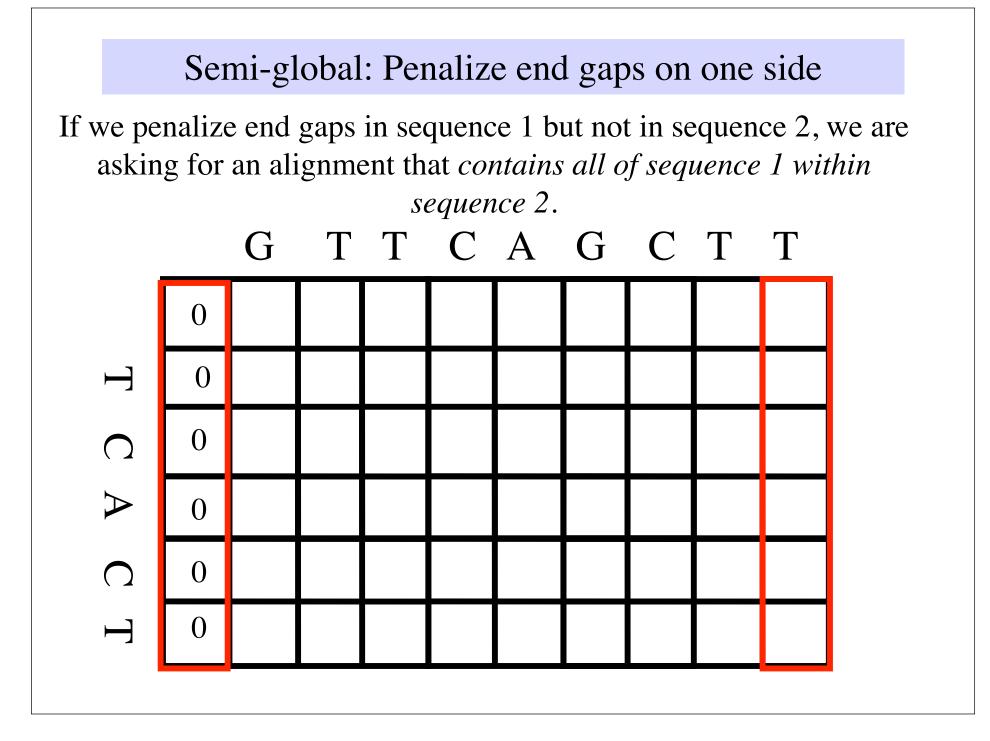


Since truncation is common in evolution, it makes sense to NOT penalize end gaps, or penalize them less than internal gaps.





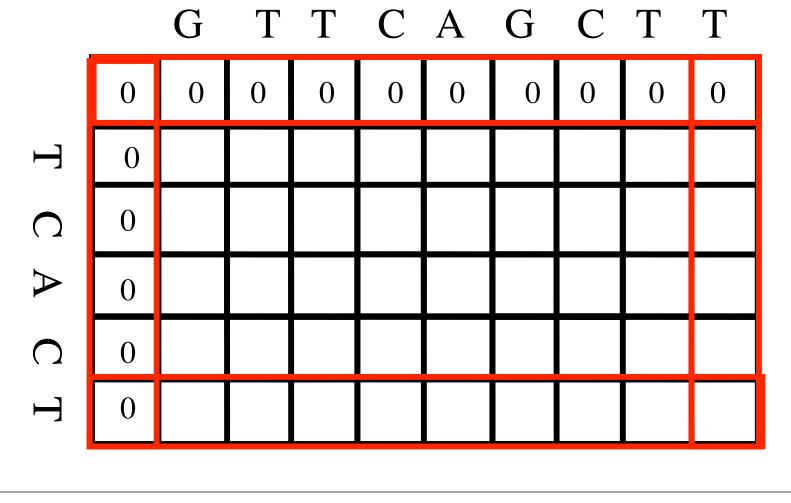


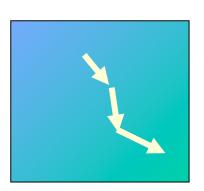


Semi-global: no end gaps

If we penalize end gaps in neither sequence, we are asking for the best alignment that contains at least two of the 4 termini.

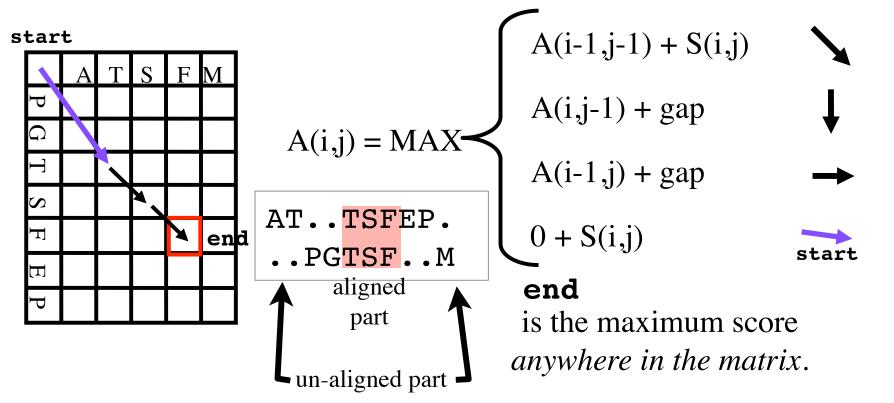
Good for identifying terminal domains in two multi-domain proteins.

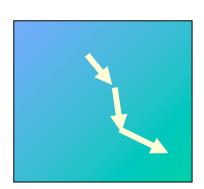




Local Alignment

A local alignment can start anywhere and end anywhere in the alignment matrix.



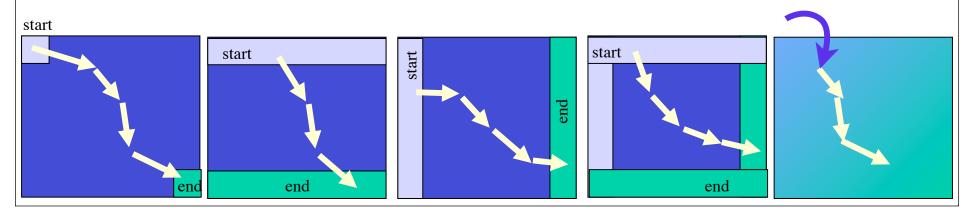


Local Alignment

- Asks for longest contiguous similarity between two sequences.
- Worst local alignment score is zero (0)
 "no alignment"
- Usually more appropriate than global or semi-global.
- Local alignment is always used for database searches.
- Local alignment scores have a theoretical distribution, used to obtain "e-values"

Global, semi-global, and local alignment

- The choice of alignment method makes a statement about how the sequences are related. Was one sequence inserted into the other?
- •Global alignment (end gaps) requires that all 4 termini are counted. In general, the two sequences are about the same length.
- •Semi-global (no end gaps in 1 or both seqs) requires that one of the two sequences be completely contained in the other or that 2 or the 4 the termini be included.
- •Local alignment finds subsequences in both. Does not require that the termini be included in the alignment.



Which alignment is intuitively better?

AGGCTACT~T~TCA GGCTACTATATCA

AGGCTACTTT~~CA

GGCTACTATATCA

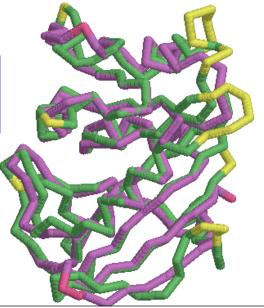
Structure-based alignments are the "gold standard"

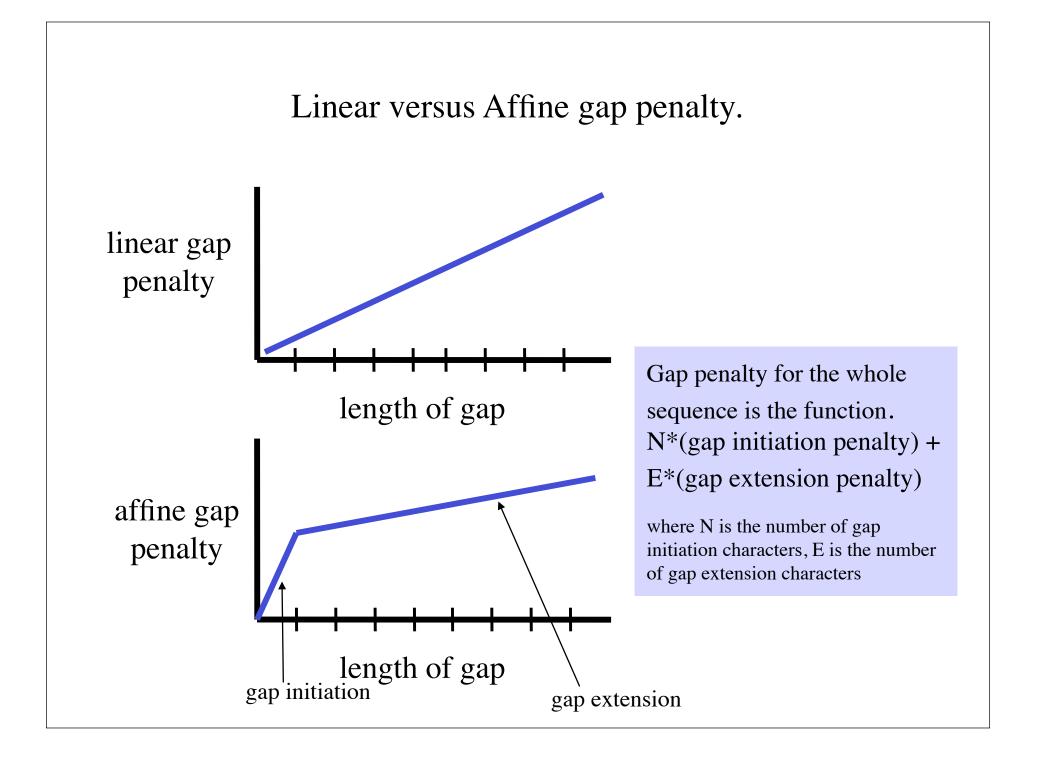
A structure-based alignment is a sequence alignment that comes from a protein structure superposition.

2DRC:A	1/2	MISLIAALAVDRVIGMENAM-PFNLPADLAWFKRNTLDKPVIMGRHTWESIG-
1DRF:_	3/4	SLNCIVAVSQNMGIGKNGDL <mark>P</mark> WPPLRNEFRYFQRMTT <mark>TSSVEGK</mark> QNLVIMGKKTWFSIP <mark>E</mark>
2DRC:A	52/53	RPLPGRKNIILSSQPGTDDRVTWVKSVDEAIAACGDVPEIMVIGGGRVYE
1DRF:_	63/64	<mark>KN</mark> RPLKGRINLVLSREL <mark>KE</mark> PPQGAHFLSRSLDDALKLTE <mark>QPELAN</mark> KVDMVWIVGGSSVYK
2DRC:A	102/103	QFLPKAQKLYLTHIDAEVEGDTHFPDYEPDDWESVFSEFHDA <mark>DAQ</mark> NSHSYCF
1DRF:_	123/124	EAMNH <mark>PG</mark> HLKLFVTRIMQDFESDTFFPEIDLEKYKLLP <mark>EYPGVL</mark> SDVQEEKGIKYKF
2DRC:A	154/155	EILERR
1DRF:_	180/181	EVYEKN

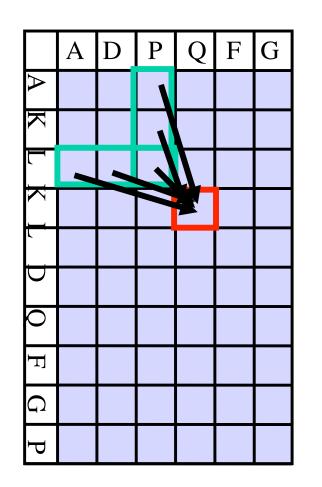
What do you see? Lots of mismatches (id=38%), few gaps (8), gaps are usually *long* (1-7).

Two similar structures may be superimposed. The parts that overlay well are the matches (purple and green), and the parts that do not overlay well are the insertions (yellow and red). Aligned positions have similar chemical 3D environment





Affine gap Dynamic Programming algorithm using variable length arrows



$$\begin{split} S_{i,j} &= \max_{n} \{ S_{i-1,j-1} + s(i,j), \\ S_{i-1-n,j-1} + s(i,j) - g_{init} - (n-1) g_{ext}, \\ S_{i-1,j-1-n} + s(i,j) - g_{init} - (n-1) g_{ext} \} \end{split}$$

...where s(i,j) is the substitution score, *n* is the length of the gap, g_{init} is the gap initiation penalty, and g_{ext} is the gap extension penalty.

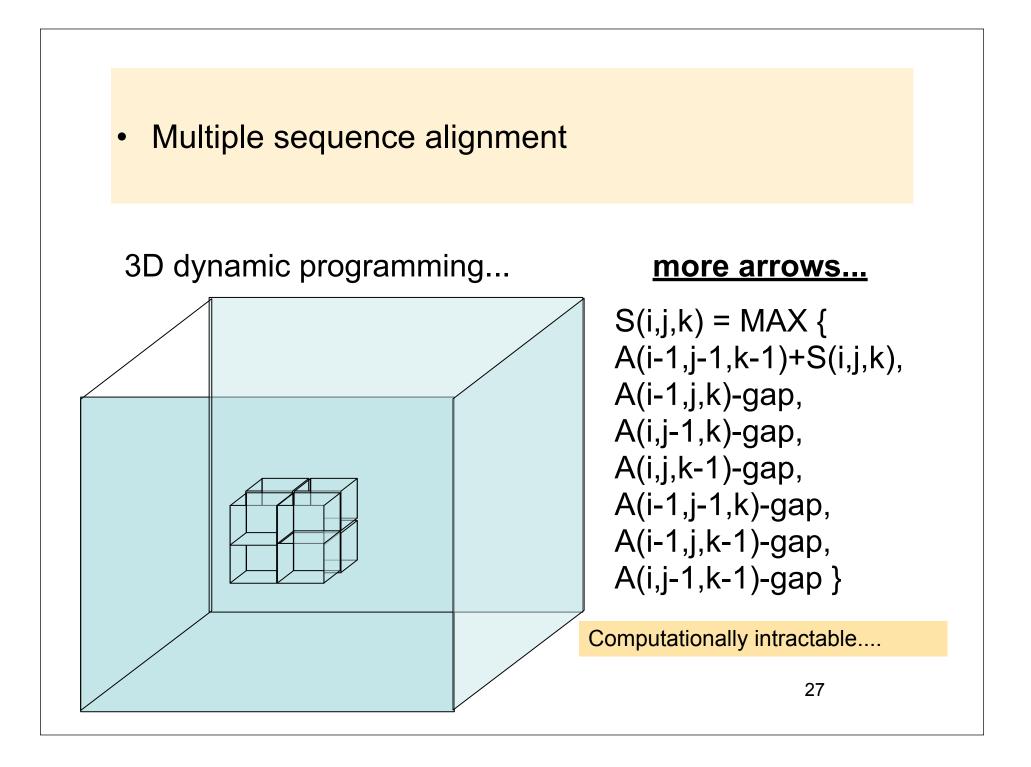
Notes: All arrows end in match. Gap-to-gap not possible. Local or semi-global only. End-gaps not scored. Arrows still translate to an alignment. Still optimal.

In class exercise: do an alignment using BLAST

- In a browser, goto to NCBI BLAST
- Protein blast.
- Align two or more sequences.
- Query: 4DDR_A
- Subjects: 2DRC_A, CAD25017
- BLAST
- Other reports: Multiple alignment. (Cobalt)
- View format: expanded, Conservation setting: Identity
- Where is the conserved region of this enzyme?

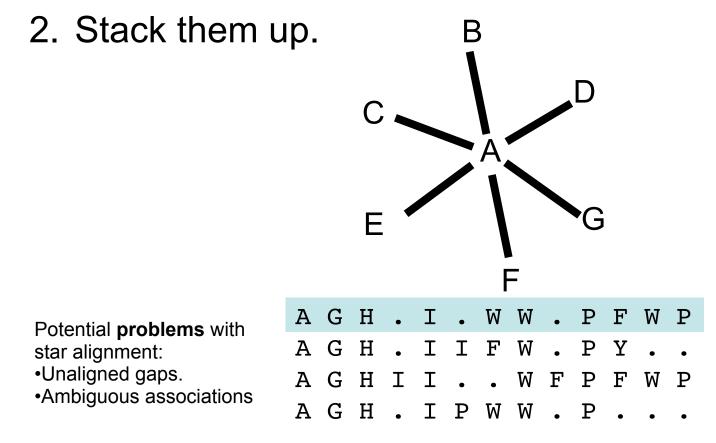
Some things to ponder

- *How does scoring approximate the evolutionary distance*
- How could you locate domain boundaries using Semiglobal alignment
- *How is dynamic programming different for local alignment?*
- Is the affine gap penalty more biologically relevant than a linear gap penalty? Why?
- Why are structure-based alignments considered the gold standard of sequence alignment?
- What does it mean for a deletion to follow immediately after an insertion, evolutionarilly? Structurally?



Multiple sequence alignment -- Star method

1. Align all sequences to one sequence.



Each pairwise alignment by itself looks fine, but when you stack them up, you see disagreements.

BLAST "query-anchored" alignments are star alignments

Query	61	TKI-SFKL-GEEFDETADNRK	80
<u>YP_003434682</u>	89	VRI-DFRV-GDAENLPFDD-EEFDAAV	112
<u>YP_004597294</u>	254	TRI-AFQN-GEETFDEST	269
<u>YP_003816569</u>	158	VAI-SASARGRPFRG-LTAAGKK	178
YP_003649443	73	TLL-SFKL-ALL-MAYASLLTGEDYRR	94
09027331	158	VTV-SFTT-NEQLNETVD	174
<u>YP_003402603</u>	247	TRI-AFQN-GEDTFDEST	262
TP_002841990	9	SFQP-EEEYVY-LTYSLKNNKK	28
<u>YP_875786</u>	521	IIH-SFSL-GTAFDETA	536
<u>ZP_09729649</u>	44	FKP-ELRV-EVEFPEQSEEMKK	63
<u>NP_280861</u>	737	GAL-SVPV-GMFGA-PDADTLT	755
<u>003481933</u>	236	AVA-WFLD-GTR	246
YP_875815	2156	AIY-QYAL-SAPFDLTSADVIS	2175
YP_876418	1736	SII-QYLL-TDSFDTSTASNLTLRR	1758
<u>YP_657166</u>	80	YDE-RVQL-PTRVDEHSAD	96
TF_004290878	444	GTV-TLQT	456
YP_006401548	88	IKV-VIRD-GEYYYVTKGDNNS	107
TP 003404280	759	NGT-VTDL-ELEFDSPLSENAT	778
YP_006351065	31	R	31
YP_001030502	56	GLK-SGKIGKHQQIL-GRELDLDILGNIDAIEAK	87
YP_002836967	25	NNKK	28
TP_004289429	173	AKI-EYLH-GEKINEN	186
YP_005645482	102	EKY-SFPS-GEKFRKVNLVSRK	121
TP 002566353	516	AAD-RPAD-APEAYIDNNASQNEA	537
<u>005380088</u>	108	SFRL-VMEVDARPDYNRK	124
<u>NP_344018</u>	11	ĸ	11
NP_634820	211	KEV-AL	215
<u>YP_003401284</u>	100	EEI-CFKI-AEEIVEGKFGKFDR-ETALDKA	127
ZP_09950113	93	TTL-TVTL-DATVTLSDTDT	110
NP_632294	14	DFEE-ITADAGS	24
YP_137647	723	REI-AYETADRN	736
<u>YP_004341996</u>	48	EIEDGAE	55
YP_003860131	68	ATI-NIPT-MEQIDVVYSVGSVSGRE	91
YP_003668585	166	NGV-RFVL-GEKVVN-IVTRDKQ	185
YP_876967	360	TVV-RYDL-DEDTVLDTSTSPNRR	381
	447	YKL-GYRD-GDDY	457
			0.05

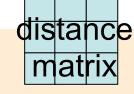
29

How can you tell? Very gappy.

Multiple sequence alignment --Progressive method

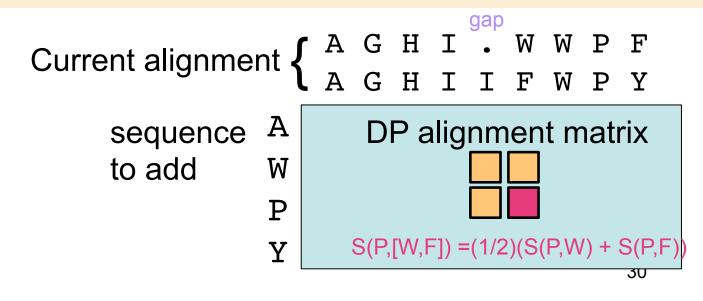
Method for progressive alignment

1. Align all pairs. Save scores in a



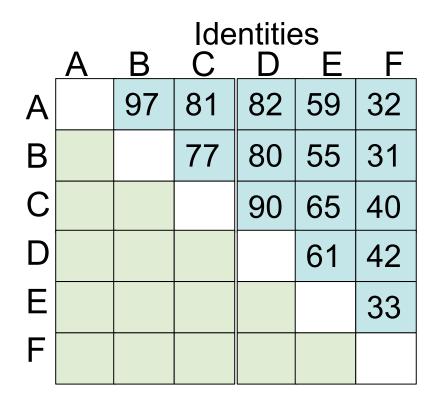
guide tree

- 2. Pairwise align two most similar.
- 3. Align the next two most similar sequence. Etc.
- 4. Add sequences until all sequences are aligned



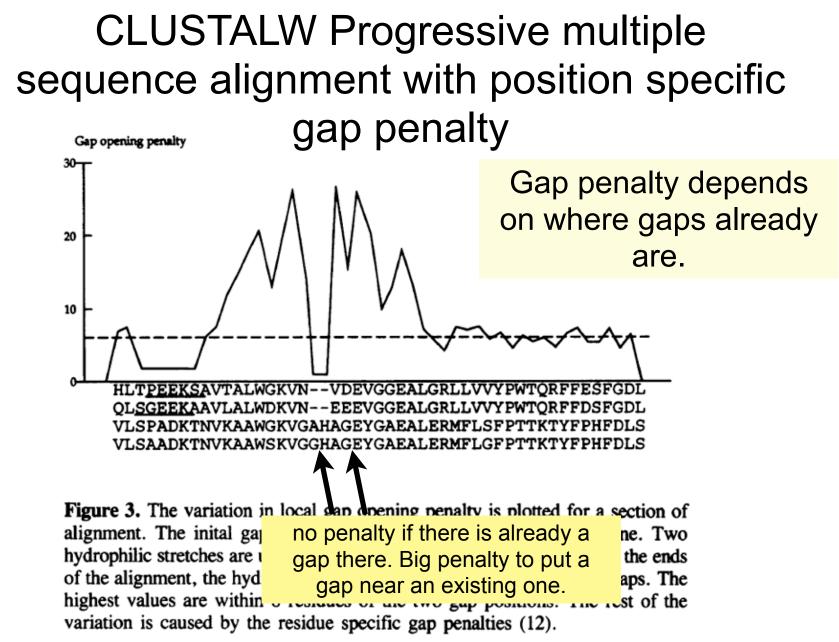
In class: progressive alignment Making a guide tree

Neighbor-joining algorithm:



ABCDEF

Draw guide tree here



J۲

Substitution matrices

- •Used to score aligned positions, usually of amino acids.
- •Expressed as the *log-likelihood ratio of mutation* (or *log-odds ratio*)
- •Derived from multiple sequence alignments

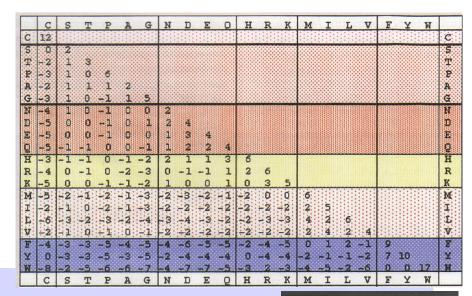
Most commonly used: PAM and BLOSUM

•PAM = percent accepted mutations (Dayhoff)

•BLOSUM = **Blo**cks **substitution matrix** (Henikoff)

PAM

M Dayhoff, 1978



•Stands for Percent Accepted Mutations

•The PAM1 matrix is made from alignments with 1% changes (99% identities).

•To get the relative frequency of each type of mutation, we count the times it was observed ^{Marg} in a database over a large set of sequence alignments.

•Based on global alignments

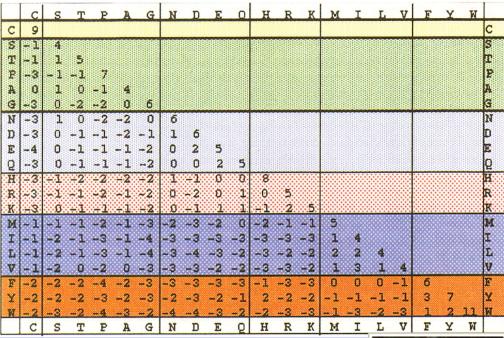


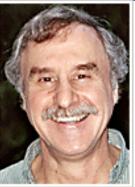
Margaret Oakley Dayhoff

BLOSUM Henikoff & Henikoff, 1992

•Based on database of ungapped local alignments (BLOCKS database)

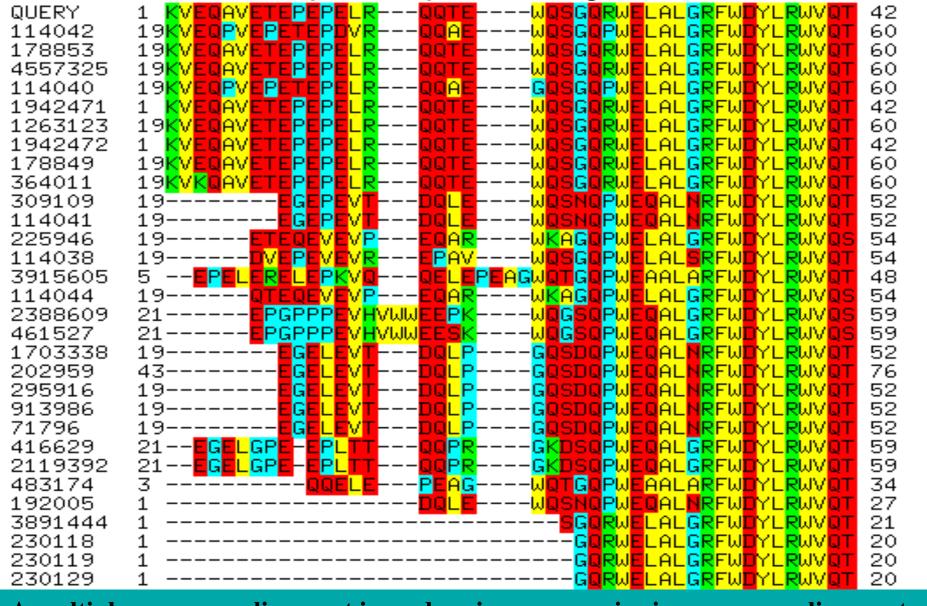
•BLOSUM number indicates the percent identity level of sequences in the alignment. For example, for BLOSUM62 sequences with approximately 62% identity were counted.





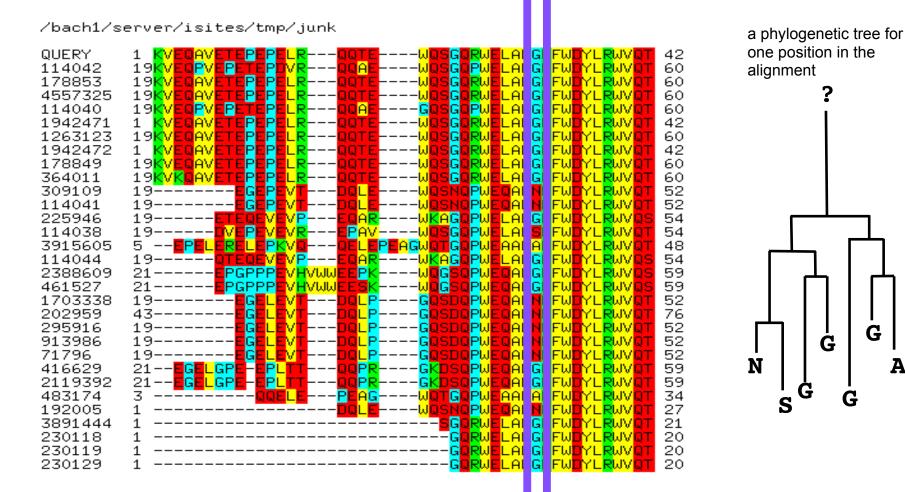
Steven Henikoff

Multiple Sequence Alignment



A multiple sequence alignment is made using many pairwise sequence alignments

Columns in a MSA have a common evolutionary history

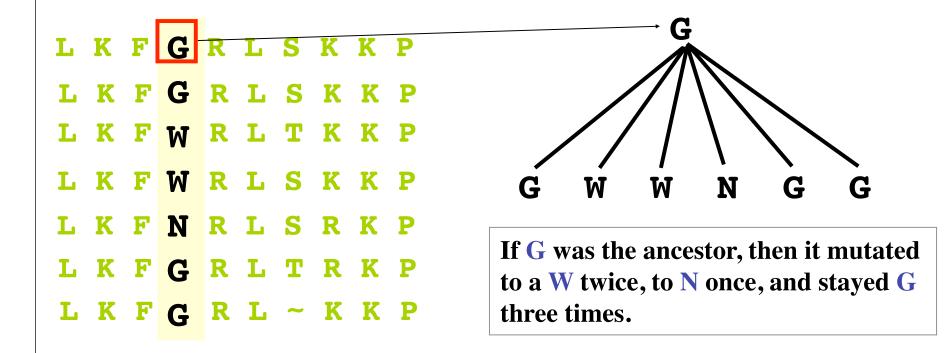


Α

By *aligning the sequences*, we are asserting that the aligned residues in each column had a common ancestor.

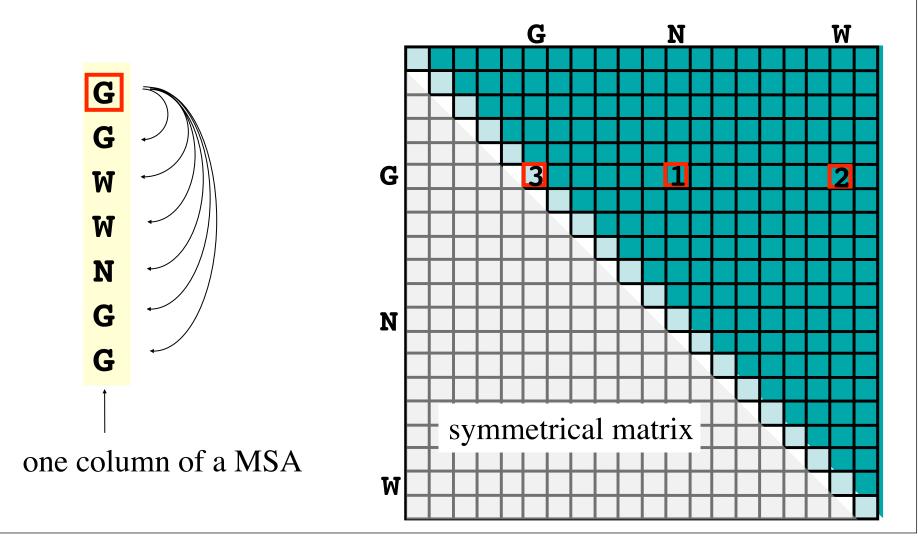
Counting mutations without knowing ancestral sequences

Naíve way: Assume *any* of the characters could be the ancestral one. Assume equal distance to the ancestor from each taxon.



Summing the substitution counts

We assume the ancestor is one of the observed amino acids, but we don't know which, so we try them all.



Substitution scores are expressed as log odds ratios

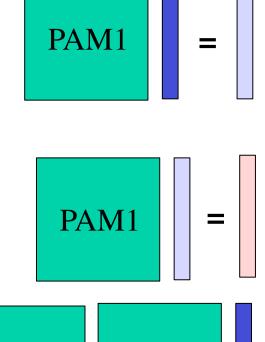
$\log \text{ odds ratio} = \log_2(\text{observed/expected})$

	C	S	т	P	A	G	N	ם	E	0	H	R	K	M	I	L	v	F	Y	W	
С	9																				С
5	-1	4					1000						101011								13
T	-1	1	5																		T
P	-3	~ 1	-1	7																	P
A	0	1	Ø	-1	4																AG
G	-3	Q	-2	-2	0	6															ŝ
N	-3	1	0	-2	-2	0	6														N
D	-3	O	-1	-1	-2	-1	1	6													D
E	-4	0	-1	-1	-1	-2	0	2	5												E
Q	-3	Ø	-1	-1	-1	-2	Ø	0	2	5											Q
H	-3	$-\mathbf{I}$	-2	-2	-2	-2	1	-1	Ø	0	8										HRK
R	~3	÷1	-1	-2	-1	~2	O.	-2	Ð	1	0	5									R
K	-3	Ø	-1	-1	-1	-2	0	-1	2	1	-1	2	5								
M	~1	-1	~1	-2	-1	~3	-2	-3	-2	0	-2	-1	~1	5							M
I	-1	-2	-1	-3	-1	-4	-3	-3	-3	-3	-3	-3	-3	1	4						I
L	-1	~2	-1	-3	-1	-4	~3	-4	~3	-2	-3	~2	-2	2	2	4					L
V	-1	-2	0	-2	0	-3	-3	-3	-2	-2	-3	-3	-2	1	3	1	4				V
F	-2	-2	-2	-4	-2	-3	-3	-3	-3	-3	-1	-3	-3	0	0	0	-1	6			F
Y	-2	-2	-2	-3	-2	-3	-2	-3	-2	-1	2	-2	-2	-1	-1	-1	-1	3	7		X
W	7	-3	-2	-4	-3	-2	-4	-4	-3	-2	-2	-3	-3	-1	-3	-2	-3	1	2	11	W
	С	S	T	P	A	G	N	D	E	Q	H	R	K	M	I	L	V	F	Y	W	

PAM assumes Markovian evolution

is just

Start with one sequence. One position. Say Gly. **Wait 1 million years**. What amino acids are now found at that position?



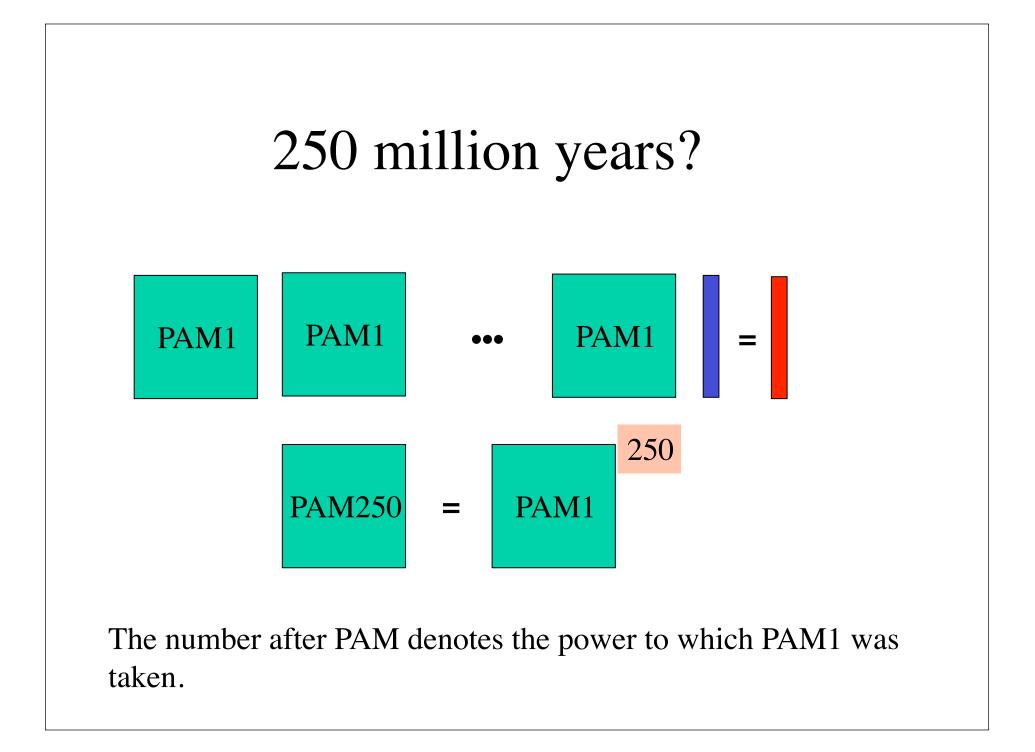
PAM1

PAM1

But,...

Wait another million years.





• NOTE OF CLARIFICATION:

- PAM does **not** stand for Plus A Million years (or anything like that). It stands for Percent Accepted <u>Mutations</u>.
- One PAM1 unit does **not** correspond to 1 million years of evolution. There is no timescale associated with PAM.
- PAM1 corresponds to 1% mutations. (or 99% identity). The timescale depends on the species.

Protein versus DNA alignments

Are protein alignment better?

- Protein alphabet = 20, DNA alphabet = 4.
 - Protein alignment is more informative
 - Less chance of homoplasy with proteins.
 - Homology detectable at greater edit distance
 - Protein alignment more informative
- Better Gold Standard alignments are available for proteins.
 - Better statistics from G.S. alignments.
- On the other hand, DNA alignments are more sensitive to short evolutionary distances. 44